

# Mode of Action of MCB3681 in *Staphylococcus aureus*-A Proteomic Study

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## Abstract

**Background:** MCB3681, a novel quinolonyl-oxazolidinone antibacterial, is active against quinolone- and/or linezolid-resistant Gram-positive bacteria. Thus, MCB3681 may interact with other targets than quinolones and oxazolidinones.

**Methods and Findings:** Protein expression in *S. aureus* following short-term exposure to MCB3681 was analyzed using 2D-gel electrophoresis and MALDI-TOF-MS/MS. Synthesis of 13 and 16 proteins was induced or repressed, respectively. Among the induced proteins are four ribosomal proteins. Repressed are proteins from different amino acid synthesis pathways, two aminoacyl-tRNA-synthetases, and methicillin-resistance-factor-protein FemB.

**Conclusion:** Our findings demonstrate that the impact of MCB3681 on the proteome signature of treated *S. aureus* cells is different from that of either ciprofloxacin or linezolid.

**Keywords:** Quinolone; *Staphylococcus aureus*; 2D-Gel electrophoresis; Protein expression; Proteome

## Introduction

Proteomics is a powerful tool to study differential protein expression of bacteria in response to antibiotics. Studying changes of the bacterial proteome in response to antibiotics allows conclusions about the mode of action and the targets of the antibiotic [1-5].

MCB3681 is a novel small molecule with structural elements of an oxazolidinone and a quinolone, showing good activity against Gram-positive bacteria irrespective of whether the isolates are quinolone- and/or linezolid-resistant [6,7]. The minimal inhibition concentrations (MICs) of MCB3681 are significantly lower than those of the 1:1 combination of ciprofloxacin and linezolid, thus demonstrating that its intrinsic activity is qualitatively and quantitatively different from oxazolidinones and quinolones. From MCB3681's activity against

ciprofloxacin and linezolid double-resistant mutants it may be hypothesized, that MCB3681 interacts with targets other than the known quinolone and oxazolidinone targets [6,7]. Therefore, the aim of this study was to examine if MCB3681 may affect expression of proteins other than those affected by ciprofloxacin or linezolid.

## Methods

The quality-control strain *S. aureus* ATCC 29213 and the test strain *S. aureus* RN1HG001 were grown aerobically under vigorous agitation at 37°C in a defined medium [8,9]. Strain RN1HG001 was used because it is susceptible to most antibiotics. In strain RN1HG001 the *rsbU* gene of *S. aureus* RN1 (NCTC8325), a strain widely used in research projects, is repaired [9]. MICs were determined in duplicate according to guidelines of the Clinical and Laboratory Standards Institute (CLSI) [10].

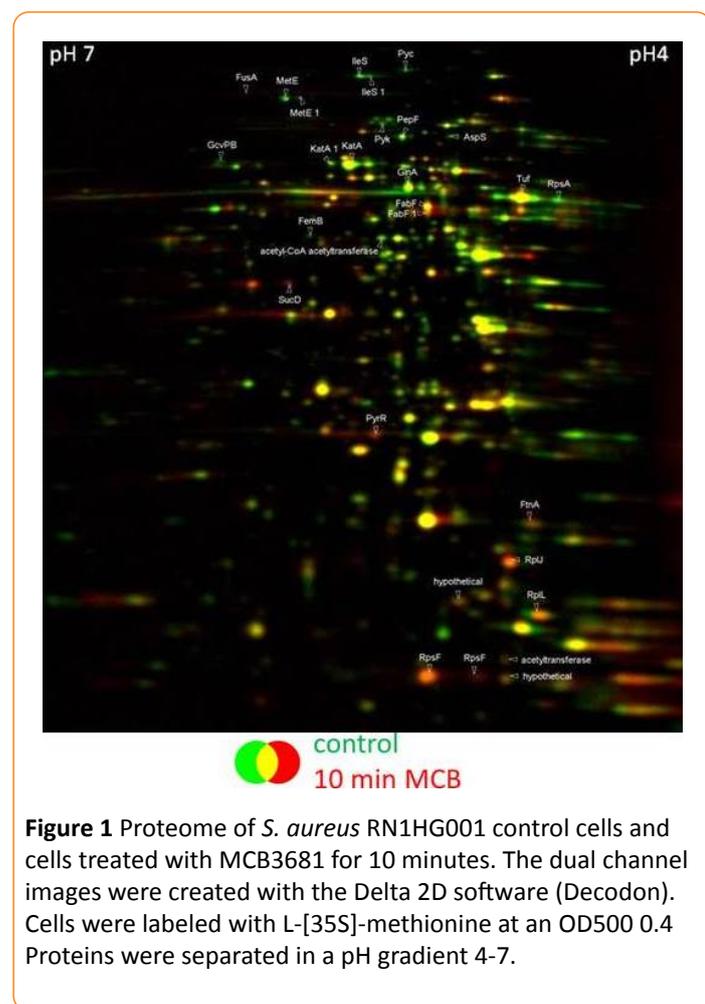
Proteins were pulse labeled with L-[35S]-methionine according to Engelmann et al. [11]. Cells at an OD500 0.4 were exposed to either 4 or 20 mg/L MCB3681 for 5 min before adding labeled methionine (3.7 MBq). Incorporation was stopped 5 min later by adding excess unlabeled methionine and chloramphenicol. Preparation of cytosolic protein extracts, protein quantification and measurement of methionine incorporation were done according to Reiss et al. [12], except that for disruption of bacteria a Precellys 24 homogenizer (Peqlab) was used twice for 30 s at 6800 rpm, separated by 5 min incubation on ice. Proteins for preparative 2D gels were obtained from bacteria exposed at an OD500 0.4 to 4 mg/L MCB3681 for 30 min.

Protein extracts (100 µg for radiolabeled samples, 300 µg for preparative gels) were separated using commercially available IPG-strips in the pH range of 4-7 [13]. Labeled gels were dried on a heated vacuum dryer followed by exposure to storage phosphor screens. Screens were scanned using a Typhoon Scanner according to Reiss et al. [12]. Preparative gels were stained and scanned as described previously [14]. For protein identification, spots were cut from preparative gels. Mass spectrometry was performed according to Wolf et al. [15]. A database including complete or partial sequences of more than 100 *S. aureus* strains available from the NCBI website was used

for searching. Search parameters were the same as described in previous studies [15]. Only proteins whose synthesis was induced or repressed at least twice in all three biological replicates were identified. Gels were analyzed using the Delta2D software (Decodon) [16].

## Results and Discussion

Both *S. aureus* strains are inhibited by 0.25 mg/L MCB3681 and concentrations up to 10 mg/L are bacteriostatic while an immediate inhibition of protein synthesis was observed at 20 mg/L (**Figure S1**). Stained gels of protein samples exposed to 20 mg/L MCB3681 resemble the controls. One structural element of MCB3681 is part of an oxazolidinone which are known to inhibit protein synthesis almost immediately, with the effect that the proteomes are “frozen in time” [17,18]. Therefore, labeling experiments were repeated with 4 mg/L MCB3681. Synthesis of most proteins is not affected by MCB3681 (**Figure 1**). Synthesis of 13 proteins is induced, and synthesis of 16 proteins is repressed. A total of 27 out of 29 protein spots with altered synthesis could be identified; all are cytosolic proteins (**Table 1 and Table S1**).



**Figure 1** Proteome of *S. aureus* RN1HG001 control cells and cells treated with MCB3681 for 10 minutes. The dual channel images were created with the Delta 2D software (Decodon). Cells were labeled with L-[35S]-methionine at an OD500 0.4. Proteins were separated in a pH gradient 4-7.

Among the induced proteins are the bifunctional protein PyrR and the ferritin family protein FtnA. FabF, 3-oxoacyl-[acyl-carrier-protein] synthase 2, occurs in two spots, only one of these spots is induced. Synthesis of four ribosomal proteins (RplL, RpsF, RplJ, RpsF) is induced, too. One ribosomal protein (RpsA) and two aminoacyl-tRNA synthetases (IleS, AspS) are synthesized at a lower level. Synthesis of ribosomal proteins is induced immediately while synthesis of most aminoacyl-tRNA-synthetases is reduced as a downstream-effect by translation inhibitors like linezolid [19] or amino-acid starvation. However, MCB3681 reduces synthesis of the isoleucyl-tRNA-synthetase and aspartyl-tRNA-synthetase instantly. This strongly reduced expression of two aminoacyl-tRNA-synthetases may represent an additional mode of action of MCB3681.

Synthesis of proteins from different amino acid synthesis pathways (GlyA, MetE, GlnA) is reduced in MCB3681 treated cells. Furthermore, exposure to MCB3681 causes a significantly reduced synthesis of methicillin-resistance factor protein FemB and of catalase, KatA, which forms three distinct spots in the gel. The reduced expression of the femB and katA gene products may be clinically relevant. Methicillin resistance factor proteins A and B are equally important for expression of methicillin-resistance [20]. Reduced expression of either of these proteins results in an increased  $\beta$ -lactam susceptibility [20]. Reduced synthesis of FemB due to exposure to MCB3681 likely contributes to its pronounced activity against MRSA. KatA on the other hand is required for nasal colonization [17]. Thus, MCB3681 may modulate adhesion of pathogens to mucosal surfaces.

Analysis of protein expression patterns does not reveal any effects of MCB3681 on DNA-replication mechanisms and the SOS-response although one structural element of MCB3681 is part of a quinolone. These findings are, nevertheless, in agreement with its mode of action. The biochemical analysis revealed that affinity of MCB3681 to topoisomerase IV is low and affinity to DNA-gyrase is 10-times lower than that of ciprofloxacin [18]. Transcriptional profiling of ciprofloxacin's effect on gene-expression revealed that only 20% of the genes involved in translation were repressed [1].

Both, the moderate effect of ciprofloxacin on the translational apparatus and the moderate affinity of MCB3681 to type II topoisomerases may explain why an effect of MCB3681 on the DNA replication machinery is not noted in this study. Likewise, the SOS-response is induced moderately by sub-inhibitory concentrations of MCB3681 whereas supra-inhibitory MCB3681 concentrations have no effect on the SOS-response [18]. The indicator strain used in this study has been exposed to supra-inhibitory MCB3681 concentrations for 10 minutes only, thus avoiding falsifying secondary effects resulting from downstream effects triggered by target inhibition. Therefore, short-term exposure to MCB3681 does not affect SOS-response.

**Table 1** MCB3681/control synthesis ratios of proteins in MCB3681 treated *S. aureus* N1HG001 cells (ratios of  $\leq 0.5$  represent a reduced synthesis of proteins, ratios of  $>2$  represent an induced synthesis of proteins).

Label	Annotated function	Ratio	Ratio	Ratio	Mean	Standard deviation
		control1_	control1_	control1_		
		vs_MCB 1	vs_MCB 2	vs_MCB 3		
<b>Induced synthesis of proteins</b>						
RpsF1	30S ribosomal protein S6	4.8	4.85	4.84	4.83	0.02
SucD	succinyl-CoA ligase [ADP-forming] subunit alpha	3.97	2.38	3.04	3.13	0.65
RpsF	30S ribosomal protein S6	3.85	4.53	3.95	4.11	0.3
RplJ	50S ribosomal protein L10	3.64	3.71	3.14	3.5	0.25
PyrR	pyrimidine operon regulatory protein	3.51	2.63	2.8	2.98	0.38
FusA	elongation factor G	3.49	3.09	4.06	3.55	0.4
FabF1	3-oxoacyl-[acyl-carrier-protein] synthase 2	2.77	2.66	2.45	2.62	0.13
RplL	50S ribosomal protein L7/L12	2.71	3.03	2.82	2.85	0.13
SAR1041/ PurQ	hypothetical/ phospho-ribosylformylglycinamide synthase 1	2.34	2.18	2.23	2.25	0.07
SACOL2532	acetyltransferase	2.19	2.39	2.26	2.28	0.08
SACOL2379	hypothetical	2.09	2.31	2.25	2.22	0.09
FtnA	ferritin	2.08	2.9	2.76	2.58	0.36
<b>Protein synthesis not affected</b>						
FabF	3-oxoacyl-[acyl-carrier-protein] synthase 2	1.23	1.39	1.46	1.36	0.1
Tuf	elongation factor Tu	1.05	1.15	1.24	1.15	0.08
<b>Reduced synthesis of proteins</b>						
SACOL0426	acetyl-CoA acetyltransferase	0.41	0.45	0.48	0.44	0.03
KatA	catalase	0.38	0.3	0.36	0.35	0.04
Pyk	pyruvate kinase	0.39	0.41	0.44	0.42	0.02
PepF	oligoendopeptidase F	0.39	0.42	0.44	0.42	0.02
MetE1	5-methyltetrahydropteroyl- triglutamate-homocysteine methyltransferase	0.38	0.3	0.51	0.4	0.09
GlyA	serine hydroxymethyltransferase	0.33	0.09	0.46	0.29	0.15
GcvPB	glycine dehydrogenase [decarboxylating] subunit 2	0.32	0.37	0.58	0.42	0.11
IleS	isoleucine-tRNA ligase	0.32	0.24	0.35	0.3	0.05
GlnA	glutamine synthetase	0.32	0.27	0.45	0.35	0.08
Pyc	pyruvate carboxylase	0.29	0.3	0.62	0.4	0.15
AspS	aspartate-tRNA ligase	0.28	0.42	0.43	0.38	0.07
IleS1	isoleucine-tRNA ligase	0.24	0.23	0.19	0.22	0.02
RpsA	30S ribosomal protein S1	0.24	0.28	0.46	0.33	0.09
MetE	5-methyltetrahydropteroyl- triglutamate-homocysteine methyltransferase	0.22	0.23	0.23	0.23	0
FemB	Aminoacyl transferase	0.09	0.33	0.42	0.28	0.14

## Conclusions

In summary, addition of MCB3681 to growing *S. aureus* cells causes a moderate reprogramming of protein synthesis with 29 protein spots exhibiting altered synthesis. In contrast, linezolid [19] and ciprofloxacin [21] provoked a much higher number of changes in gene expression. Following linezolid- and ciprofloxacin-stress, expression of 566 [19] and 235 proteins [21], respectively, is changed. Thus, the impact of MCB3681 on the proteome signature is different from that of either ciprofloxacin or linezolid.

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## Conflict of Interest

The authors declare no conflict of interest.

## References

- Freiberg C, Fischer HP, Brunner NA (2005) Discovering the mechanism of action of novel antibacterial agents through transcriptional profiling of conditional mutants. *Antimicrob Agents Chemother* 49: 749-759.
- Bandow JE, Brötz H, Leichert LI, Labischinski H, Hecker M (2003) Proteomic approach to understanding antibiotic action. *Antimicrob Agents Chemother* 47: 948-955.
- Wecke T, Mascher T (2011) Antibiotic research in the age of omics: from expression profiles to interspecies communication. *J Antimicrob Chemother* 66: 2689-2704.
- Lee CR, Lee JH, Park KS, Jeong BC, Lee SH (2015) Quantitative proteomic view associated with resistance to clinically important antibiotics in Gram-positive bacteria: a systematic review. *Front Microbiol* 6: 828.
- Fouhy F, Stanton C, Cotter PD, Hill C, Walsh F (2015) Proteomics as the final step in the functional metagenomics study of antimicrobial resistance. *Front Microbiol* 6: 172.
- Rashid MU, Dalhoff A, Weintraub A, Nord CE (2014) In vitro activity of MCB3681 against *Clostridium difficile* strains. *Anaerobe* 28: 216-219.
- Dalhoff A, Rashid MU, Kapsner T, Panagiotidis G, Weintraub A, et al. (2015) Analysis of effects of MCB3681, the antibacterially active substance of prodrug MCB3837, on human resident microflora as proof of principle. *Clin Microbiol Infect* 21: 761-764.
- Gertz S, Engelmann S, Schmid R, Ohlsen K, Hacker J, et al. (1999) Regulation of sigmaB-dependent transcription of sigB and asp23 in two different *Staphylococcus aureus* strains. *Mol Gen Genet* 261: 558-566.
- Herbert S, Ziebandt AK, Ohlsen K, Schäfer T, Hecker M, et al. (2010) Repair of global regulators in *Staphylococcus aureus* 8325 and comparative analysis with other clinical isolates. *Infect Immun* 78: 2877-2889.
- Jean BP, Franklin RC (2009) Methods for dilution antimicrobial susceptibility tests for bacteria that grow aerobically: approved standard. Clinical and Laboratory Standards Institute (CLSI), pp: 1-86
- Engelmann S, Hecker M (2008) Proteomic analysis to investigate regulatory networks in *Staphylococcus aureus*. *Methods Mol Biol* 431: 25-45.
- Reiss S, Pane-Farre J, Fuchs S, Francois P, Liebeke M, et al. (2012) Global analysis of the *Staphylococcus aureus* response to mupirocin. *Antimicrob Agents Chemother* 56: 787-804.
- Büttner K, Bernhardt J, Scharf C, Schmid R, Mäder U, et al. (2001) A comprehensive two-dimensional map of cytosolic proteins of *Bacillus subtilis*. *Electrophoresis* 22: 2908-2935.
- Voigt B, Schweder T, Becher D, Ehrenreich A, Gottschalk G, et al. (2004) A proteomic view of cell physiology of *Bacillus licheniformis*. *Proteomics* 4: 1465-1490.
- Wolf C, Hochgräfe F, Kusch H, Albrecht D, Hecker M, et al. (2008) Proteomic analysis of antioxidant strategies of *Staphylococcus aureus*: diverse responses to different oxidants. *Proteomics* 8: 3139-3153.
- Bernhardt J, Büttner K, Scharf C, Hecker M (1999) Dual channel imaging of two-dimensional electropherograms in *Bacillus subtilis*. *Electrophoresis* 20: 2225-2240.
- Cosgrove K, Coutts G, Jonsson IM, Tarkowski A, Kokai-Kun JF, et al. (2007) Catalase (KatA) and alkyl hydroperoxide reductase (AhpC) have compensatory roles in peroxide stress resistance and are required for survival, persistence, and nasal colonization in *Staphylococcus aureus*. *J bacteriol* 189: 1025-1035.
- Gray C, Moes L, Locher H, Dale G (2005) Characterization of MCB3681, a dual-action antibiotic. 45th ICAAC: Abstract No F-51.
- Bonn F, Pane-Farre J, Schlüter R, Schaffer M, Fuchs S, et al. (2016) Global analysis of the impact of linezolid onto virulence factor production in *S. aureus* USA300. *Int J Med Microbiol* 306: 131-140.
- Henze U, Sidow T, Wecke J, Labischinski H, Berger-Bächi B (1993) Influence of femB on methicillin resistance and peptidoglycan metabolism in *Staphylococcus aureus*. *J Bacteriol* 175: 1612-1620.
- Cirz RT, Jones MB, Gingles NA, Minogue TD, Jarrahi B, et al. (2007) Complete and SOS-mediated response of *Staphylococcus aureus* to the antibiotic ciprofloxacin. *J bacteriol* 189: 531-539.